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Why does epithelia display heterogeneity? Bridging physical and biological concepts

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Abstract

Technological and computational advances in the past few decades have allowed biophysicists to describe behaviour of epithelia, using mesoscale physical principles. In such description, similar to a glassy solid or dense particulate matter, epithelial cells are shown to transit from fluid-like motion to a jammed and kinetically arrested state upon crowding. This jamming transition is characterized by dynamic heterogeneity, which is revealed by fluctuations in intercellular stresses arising from multicellular cooperation. Even though recent studies are suggestive of the role of dynamic heterogeneity in tissue homeostasis and wound healing, very little is known about its physiological meaning. Meanwhile, recent studies in epithelial cell biology reveal an intrinsic cellular heterogeneity arising from variations in genome and protein expression patterns. Interestingly, such heterogeneity is also shown to be relevant in regulating tissue homeostasis. In the light of such observations, it becomes intuitive to ask how the inherent biological heterogeneity of epithelial tissues influences its physical behaviour. In this review, we attempt to bridge this gap by connecting studies describing dynamic heterogeneity in epithelial tissues with those that describe intrinsic biological heterogeneity and discuss how these might be linked in order to potentially regulate epithelial functionality.

Key words: heterogeneity, epithelial tissue, jamming, collective behaviour, extra-cellular matrix

Epithelial cells construct inner and outer linings of our organs and function as physical barriers, thus, protecting the underlying tissue from infections, dehydration, and also aiding in efficient absorption of nutrients and gases (Alberts 2008). Cells within the epithelia perform these tasks, being jammed at their place while also making sure that epithelial homeostasis is maintained, failing in which can be potentially fatal for the tissue (Macara et al. 2014). Interestingly, the same cells can unjam and flow almost like a fluid during physiological and pathological situations such as organ development, wound healing and cancer metastasis (Friedl and Gilmour 2009; Mongera et al. 2018; Park et al. 2016; Sadati et al. 2013; Scarpa and Mayor 2016). In such situations, cells, rather than moving individually, migrate as a group in various patterns (Haeger et al. 2015; Petitjean et al. 2010; Poujade et al. 2007; Rorth 2012; Tarle et al. 2015). Reductionist view holds that such cooperative cellular events are mediated at the level of cell-cell interactions where local signals are translated into physical forces (such as those generated in the cellular cytoskeleton and those exerted across cell-cell junctions), which are then translated into cell motility (Das et al. 2015; Keller 2012; Ladoux and Mège 2017; Trepap et al. 2009). Such physical forces are believed to be fundamental to biological form and function but have remained hidden until recently when experimental methods are finally making them visible (Angelini et al. 2010; Angelini et al. 2011; Edwards and Schwarz 2011; Malinverno et al. 2017; Sabass et al. 2008; Schwarz and Soine 2015; Sunyer et al. 2016; Tambe et al. 2011; Trepap and Fredberg 2011). Furthermore, recent advances in mathematical biology have also led to the development of models that can predict various parameters of epithelial behaviour in both jammed and unjammed states (Edwards and Schwarz 2011; Garcia et al. 2015; Henkes et al. 2011; Mark et al. 2010; Mehes and Vicsek 2014; Sepulveda et al. 2013; Steinberg 2007). Together, these studies have revealed unpredicted behaviour of epithelial tissues and are beginning to explain why cells jam and unjam, and how collective cell behaviour is orchestrated. Since excellent reviews have been written on the topic (Friedl and Gilmour 2009; Haeger et al. 2015; Merkel and Manning 2017; Park et al. 2016; Park and Fredberg 2016; Pegoraro et al. 2016; Sadati et al. 2013), we will only briefly describe the heterogeneous nature of the jamming transition from the physical perspective and then discuss its implications in regulating epithelial functionality while also taking into account the inherent biological heterogeneity present within the epithelium.

Jamming transition and dynamic heterogeneity. Ongoing cell divisions, apoptosis and cell mingling make the epithelia a highly dynamic place (Al-Hussaini et al. 2016; Christ et al. 1990; Gardner 1986; Macara et al. 2014). Interestingly, monolayer stress profiles of such epithelial layers reveal dynamic heterogeneity, with intercellular stress displaying stochasticity in space and time, meaning that stress is tied neither to any particular position nor to any particular cell within the monolayer (Angelini et al. 2010; Angelini et al. 2011; Garrahan 2011; Tambe et al. 2011). Topography of these intercellular forces, at any given instant, can be compared with a rugged landscape, similar to that of a mountain range, where peaks arise from cooperation between tens of cells pulling together (Tambe et al. 2011) (Fig. 1a). Interestingly, cell density also plays a key role in regulating dynamic heterogeneity i.e. when cells start to crowd, their movement becomes arrested and zones of cooperativity grow bigger (Angelini et al. 2011). Such a scenario is intriguingly analogous to glass transition within a supercooled fluid or dense particulate matter in which a non-equilibrium jammed state is reached by cooling, crowding or by decreasing applied load (Debenedetti and Stillinger 2001; Mattsson et al. 2009; Mayer et al. 2008; Nagel 1998; Trappe et al. 2001). Hallmarks similar to glass transition (spontaneous intermittent fluctuations, dynamic heterogeneity, cooperativity, and kinetic arrest) are observed by epithelial cell monolayer, wherein, the dynamical arrest is caused

upon crowding and depends upon parameters such as active motility, cellular forces, cell shape and applied stress. When these parameters are comprehended in a jamming phase diagram (Nagel 1998; Sadati et al. 2013; Trappe et al. 2001), predictions on epithelial physical behaviour can be made. For instance, as intercellular adhesion or crowding progressively increases, cell motility and rearrangement would become rare and therefore, cooperativity would increase, leading to a topologically frozen epithelium (Sadati et al. 2013). Subsequently, then, the question is what the extension of jamming at homeostasis should be that allows the epithelia to achieve their vital physiological functions such as regulating homeostasis and orchestrating collective cell migration.

Physiological relevance of heterogeneity. The ability of epithelial cells to dynamically remodel their surroundings as well as their own cytoskeleton in response to external cues such as damage or mechanical stresses is known to provide a mechanical resilience to epithelial tissues (Khalilgharibi et al. 2019; Treppe and Sahai 2018). Recent studies are suggestive of the hypothesis that, by maintaining a striking balance between jammed and unjammed phases, the epithelial monolayer might have evolved to attain such resilience, by virtue of which, it can efficiently undergo switch-like changes required for physiological functions (Park et al. 2015; Sadati et al. 2014; Saw et al. 2017; Vishwakarma et al. 2018). For instance, a recent study demonstrates that cooperative forces owing to dynamic heterogeneity control the selection as well as frequency of leader cells which guide collective migration during wound healing (Vishwakarma et al. 2018). Another study demonstrates that hot spots of compressive stresses within the epithelial monolayer induce topological defects that subsequently lead to local cell extrusion (Saw et al. 2017). Since hot spots of compressive stresses build up regions of multicellular cooperation (Tambe et al. 2011) which show density dependence (Angelini et al. 2011), efficient cell extrusion for regulating tissue homeostasis would intuitively require the right extent of cell packing. Such extrusion events are important, not only due to their relevance in regulating cell density during epithelial homeostasis (Fadul and Rosenblatt 2018; Gudipaty et al. 2018) but also in removing aberrant or tumour cells via a mechanism described as cell competition, by virtue of which, epithelia gains the ability to defend itself against cancer (Kajita and Fujita 2015; Wagstaff et al. 2013). Understanding physiological relevance and extent of jamming in epithelia becomes even more important in tissues that are naturally subjected to elevated levels of stress, such as lung epithelium which goes through cyclic breathing stress and, therefore, tissue plasticity plays an important role in maintaining its integrity, especially during lung injury (Frank and Matthay 2003).

The physical heterogeneity described above is most likely to be influenced by the existing innate biological heterogeneities in the epithelia which are associated with variations in genome or protein expression patterns (Fig. 1b). A known outcome of this genetic variability is somatic mosaicism that leads to the presence of multiple cell clones within an adult tissue. Somatic mosaicism can originate from epigenetics events (Rakyan et al. 2002; Sutherland et al. 2000) such as, for instance, the inactivation of one of the X-chromosomes in females (Rakyan et al. 2002) or from mobile DNA elements such as retrotransposons (Beck et al. 2011; De 2011). A classic example of somatic mosaicism can be observed in the skin with the presence of mosaicisms in the pigmentation known as *café-au-lait* spots (De 2011; Rawles 1947). In addition to these genetic differences, differential regulation of proteins expression induced by external cues, such as extracellular matrix (ECM), can also create cellular heterogeneity within the epithelial layer (Fig. 1b). The importance of such heterogeneity in regulating tissue homeostasis has been shown in the basal layer of esophageal epithelium containing stem cells responsible for

1 tissue renewal (DeWard et al. 2014). It has been shown that, in this layer, population of stem cells has
2 heterogenous proliferation rates which are distinguishable by the expression of specific cell-surface markers such
3 as the laminin receptor integrin $\alpha 6 \beta 4$. Here, the involvement of laminins, major components of extra-cellular
4 matrix, suggests the importance of cell-ECM adhesion in maintaining cellular heterogeneity, and subsequently in
5 regulating tissue homeostasis (DeWard et al. 2014). Interestingly, cellular heterogeneity dictated by differential
6 laminin expression has also been shown to be involved in regulating functionality of endothelial cells. For
7 example, the extracellular matrix of endothelium in postcapillary venules consists of areas of high and low
8 expression of the laminin 511 isoform as compared to the capillaries where the expression of laminin 511 is
9 homogeneous (Di Russo et al. 2017; Sixt et al. 2001). Such differential distribution of laminin controls endothelial
10 cell junction tightness, thereby dictating the location of leucocytes extravasation through the blood-brain barrier
11 which occurs only in low laminin 511 regions (Sixt et al. 2001; Song et al. 2017). In addition to the biochemical
12 composition of ECM, its topography has also been shown to control the heterogeneity of epithelial cells. Recently,
13 an elegant experimental setting using undulated elastomer surfaces revealed the effect of ECM topography on
14 heterogeneity of keratinocytes (Mobasserri et al. 2019). After seeding primary keratinocyte on the surfaces, the
15 monolayer assembled within a range of cellular stiffness, cell-cell adhesion forces and acto-myosin contractility
16 levels. The results provided new insights into the possible heterogenous control of keratinocytes proliferation
17 rates by the topography of the dermal ECM during ageing and inflammation (Mobasserri et al. 2019). Differential
18 ECM expression also impacts on the aetiology of retinal degenerative disease, i.e. age-related macular
19 degeneration. The early stage of the disease is characterized by high level of ECM accumulation known as *drusen*
20 that occurs between the retinal pigment epithelium and the underlying Bruch's membrane (Coleman et al. 2008).
21 Drusen formation is a common age effect, but only the accumulation of high number of large drusen ($> 63 \mu\text{m}$ in
22 diameter) correlates with epithelium degeneration and photoreceptor detachment (Coleman et al. 2008). Since the
23 retina pigment epithelium presents a very high heterogeneity in cell shape (Fig. 2), protein synthesis and granules
24 accumulation, it is tempting to speculate that this diversity of cell shape might also correspond to high
25 heterogeneity in monolayer tensions and, therefore, might control drusen formation and their growth.

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Conclusion. Even though physical and biological heterogeneities are currently known to be distinct, they are
likely to be interactive and interdependent. Local cellular heterogeneity might influence the mechanical properties
of epithelia, its ability to transduce forces and, hence, the nature of physical heterogeneity. Recent technological
advancements in biophysics, cell biology and mathematical biology has now made it possible to analyze the
physics and biology of the epithelia within the same framework. Such approach allows us to attain a more
comprehensive understanding on epithelial physiology and would subsequently require devising new treatment
strategies for epithelia degenerative diseases

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Conflict of Interest: Authors declare no conflict of interest.

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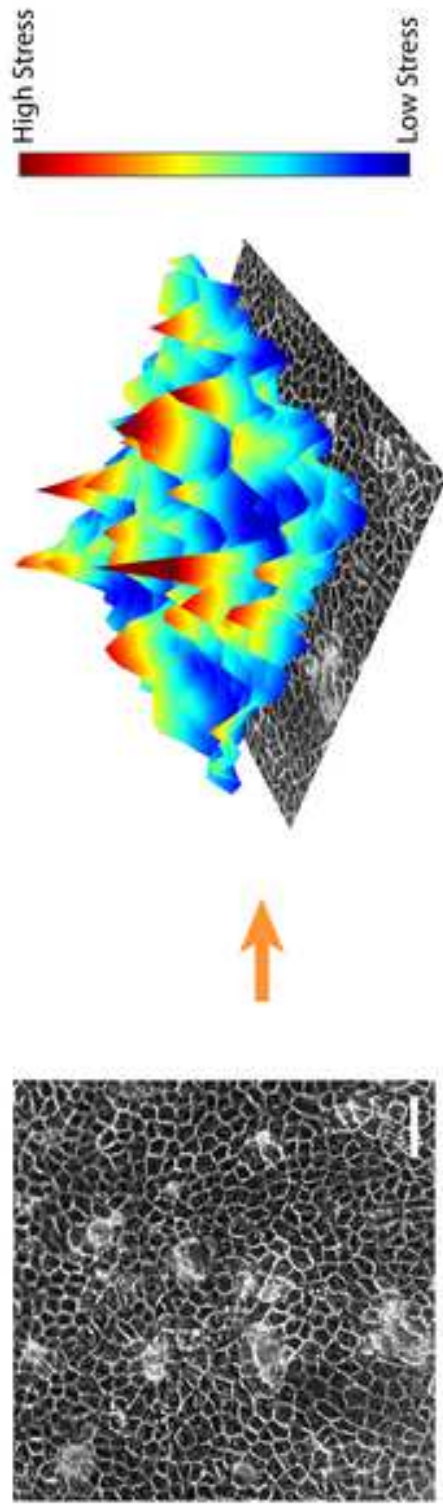
References

- Al-Hussaini H, Kilarkaje N, Shahabi G, Al-Mulla F (2016) Proliferation and Migration of Peripheral Retinal Pigment Epithelial Cells Are Associated with the Upregulation of Wntless-Related Integration and Bone Morphogenetic Protein Signaling in Dark Agouti Rats *Medical Principles and Practice* 25:408-416 doi:10.1159/000446480
- Alberts B (2008) *Molecular biology of the cell*. Reference edition. 5th edn. Garland Science, New York
- Angelini TE, Hannezo E, Trepat X, Fredberg JJ, Weitz DA (2010) Cell migration driven by cooperative substrate deformation patterns *Phys Rev Lett* 104:168104 doi:10.1103/PhysRevLett.104.168104
- Angelini TE, Hannezo E, Trepat X, Marquez M, Fredberg JJ, Weitz DA (2011) Glass-like dynamics of collective cell migration *Proc Natl Acad Sci U S A* 108:4714-4719 doi:10.1073/pnas.1010059108
- Beck CR, Garcia-Perez J, Badge RM, Moran JV (2011) LINE-1 Elements in Structural Variation and Disease *Annual Review of Genomics and Human Genetics* 12:187-215 doi:10.1146/annurev-genom-082509-141802
- Christ B, Poelmann RE, Mentink MMT, Groot G-dAC (1990) Vascular endothelial cells migrate centripetally within embryonic arteries *Anatomy and Embryology* 181:333-339 doi:10.1007/BF00186905
- Coleman HR, Chan C-C, Ferris FL, Chew EY (2008) Age-related macular degeneration *The Lancet* 372:1835-1845 doi:10.1016/S0140-6736(08)61759-6
- Das T, Safferling K, Rausch S, Grabe N, Boehm H, Spatz JP (2015) A molecular mechanotransduction pathway regulates collective migration of epithelial cells *Nat Cell Biol* 17:276-287 doi:10.1038/ncb3115
- De S (2011) Somatic mosaicism in healthy human tissues *Trends in Genetics* 27:217-223 doi:10.1016/j.tig.2011.03.002
- Debenedetti PG, Stillinger FH (2001) Supercooled liquids and the glass transition *Nature* 410:259-267 doi:10.1038/35065704
- DeWard AD, Cramer J, Lagasse E (2014) Cellular Heterogeneity in the Mouse Esophagus Implicates the Presence of a Nonquiescent Epithelial Stem Cell Population *Cell Reports* 9:701-711 doi:10.1016/j.celrep.2014.09.027
- Di Russo J et al. (2017) Vascular laminins in physiology and pathology *Matrix biology : journal of the International Society for Matrix Biology* 57-58:140-148 doi:10.1016/j.matbio.2016.06.008
- Edwards CM, Schwarz US (2011) Force localization in contracting cell layers *Phys Rev Lett* 107:128101 doi:10.1103/PhysRevLett.107.128101
- Fadul J, Rosenblatt J (2018) The forces and fates of extruding cells *Current Opinion in Cell Biology* 54:66-71 doi:10.1016/j.ceb.2018.04.007
- Frank JA, Matthay MA (2003) Science review: mechanisms of ventilator-induced injury *Critical care (London, England)* 7:233-241 doi:10.1186/cc1829
- Friedl P, Gilmour D (2009) Collective cell migration in morphogenesis, regeneration and cancer *Nat Rev Mol Cell Biol* 10:445-457 doi:10.1038/nrm2720
- Garcia S, Hannezo E, Elgeti J, Joanny JF, Silberzan P, Gov NS (2015) Physics of active jamming during collective cellular motion in a monolayer *Proc Natl Acad Sci U S A* 112:15314-15319 doi:10.1073/pnas.1510973112
- Gardner RL (1986) Cell mingling during mammalian embryogenesis *J Cell Sci Suppl* 4:337-356
- Garrahan JP (2011) Dynamic heterogeneity comes to life *Proceedings of the National Academy of Sciences* 108:4701-4702 doi:10.1073/pnas.1101436108
- Gudipaty SA, Conner CM, Rosenblatt J, Montell DJ (2018) Unconventional Ways to Live and Die: Cell Death and Survival in Development, Homeostasis, and Disease *Annual review of cell and developmental biology* 34:311-332 doi:10.1146/annurev-cellbio-100616-060748
- Haeger A, Wolf K, Zegers MM, Friedl P (2015) Collective cell migration: guidance principles and hierarchies *Trends in cell biology* 25:556-566 doi:10.1016/j.tcb.2015.06.003
- Henkes S, Fily Y, Marchetti MC (2011) Active jamming: self-propelled soft particles at high density *Phys Rev E Stat Nonlin Soft Matter Phys* 84:040301 doi:10.1103/PhysRevE.84.040301
- Kajita M, Fujita Y (2015) EDAC: Epithelial defence against cancer-cell competition between normal and transformed epithelial cells in mammals *J Biochem* 158:15-23 doi:10.1093/jb/mvv050
- Keller R (2012) Developmental biology. Physical biology returns to morphogenesis *Science* 338:201-203 doi:10.1126/science.1230718
- Khalilgharibi N et al. (2019) Stress relaxation in epithelial monolayers is controlled by the actomyosin cortex *Nature Physics* 1-9 doi:10.1038/s41567-019-0516-6
- Ladoux B, Mège R-MM (2017) Mechanobiology of collective cell behaviours *Nature reviews Molecular cell biology* 18:743-757 doi:10.1038/nrm.2017.98

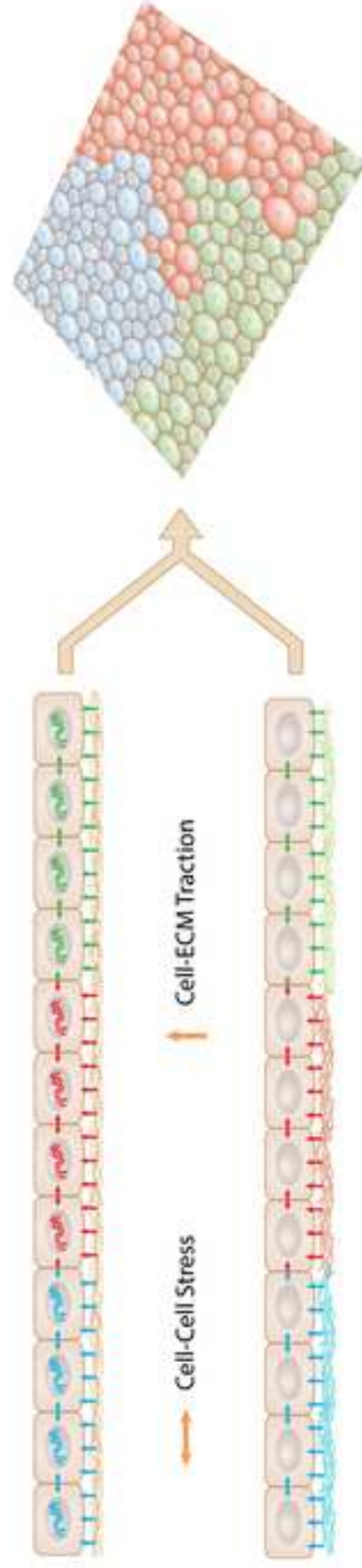
- Macara IG, Guyer R, Richardson G, Huo Y, Ahmed SM (2014) Epithelial homeostasis *Curr Biol* 24:R815-825 doi:10.1016/j.cub.2014.06.068
- Malinverno C et al. (2017) Endocytic reawakening of motility in jammed epithelia *Nat Mater* 16:587-596 doi:10.1038/nmat4848
- Mark S, Shlomovitz R, Gov NS, Poujade M, Grasland-Mongrain E, Silberzan P (2010) Physical model of the dynamic instability in an expanding cell culture *Biophys J* 98:361-370 doi:10.1016/j.bpj.2009.10.022
- Mattsson J, Wyss HM, Fernandez-Nieves A, Miyazaki K, Hu Z, Reichman DR, Weitz DA (2009) Soft colloids make strong glasses *Nature* 462:83-86 doi:10.1038/nature08457
- Mayer C et al. (2008) Asymmetric caging in soft colloidal mixtures *Nat Mater* 7:780-784 doi:10.1038/nmat2286
- Mehes E, Vicsek T (2014) Collective motion of cells: from experiments to models *Integr Biol (Camb)* 6:831-854 doi:10.1039/c4ib00115j
- Merkel M, Manning ML (2017) Using cell deformation and motion to predict forces and collective behavior in morphogenesis *Semin Cell Dev Biol* 67:161-169 doi:10.1016/j.semcdb.2016.07.029
- Mobasser SA, Zijl S, Salameti V, Walko G, Stannard A, Garcia-Manyes S, Watt FM (2019) Patterning of human epidermal stem cells on undulating elastomer substrates reflects differences in cell stiffness *Acta biomaterialia* 87:256-264 doi:10.1016/j.actbio.2019.01.063
- Mongera A et al. (2018) A fluid-to-solid jamming transition underlies vertebrate body axis elongation *Nature* 561:401-405 doi:10.1038/s41586-018-0479-2
- Nagel AJLaSR (1998) Jamming is not just cool any more *Nature* 396
- Park JA, Atia L, Mitchel JA, Fredberg JJ, Butler JP (2016) Collective migration and cell jamming in asthma, cancer and development *J Cell Sci* doi:10.1242/jcs.187922
- Park JA, Fredberg JJ (2016) Cell Jamming in the Airway Epithelium *Ann Am Thorac Soc* 13 Suppl 1:S64-67 doi:10.1513/AnnalsATS.201507-476MG
- Park JA et al. (2015) Unjamming and cell shape in the asthmatic airway epithelium *Nat Mater* 14:1040-+ doi:10.1038/NMAT4357
- Pegoraro AF, Fredberg JJ, Park JA (2016) Problems in biology with many scales of length: Cell-cell adhesion and cell jamming in collective cellular migration *Exp Cell Res* 343:54-59 doi:10.1016/j.yexcr.2015.10.036
- Petitjean L, Reffay M, Grasland-Mongrain E, Poujade M, Ladoux B, Buguin A, Silberzan P (2010) Velocity fields in a collectively migrating epithelium *Biophys J* 98:1790-1800 doi:10.1016/j.bpj.2010.01.030
- Poujade M et al. (2007) Collective migration of an epithelial monolayer in response to a model wound *Proc Natl Acad Sci U S A* 104:15988-15993 doi:10.1073/pnas.0705062104
- Rakyan VK, Blewitt ME, Druker R, Preis JJ, Whitelaw E (2002) Metastable epialleles in mammals *Trends Genet* 18:348-351
- Rawles ME (1947) Origin of pigment cells from the neural crest in the mouse embryo *Physiol Zool* 20:248-266
- Rorth P (2012) Fellow travellers: emergent properties of collective cell migration *Embo Rep* 13:984-991 doi:10.1038/Embor.2012.149
- Sabass B, Gardel ML, Waterman CM, Schwarz US (2008) High resolution traction force microscopy based on experimental and computational advances *Biophys J* 94:207-220 doi:10.1529/biophysj.107.113670
- Sadati M, Nourhani A, Fredberg JJ, Taheri Qazvini N (2014) Glass-like dynamics in the cell and in cellular collectives *Wiley Interdiscip Rev Syst Biol Med* 6:137-149 doi:10.1002/wsbm.1258
- Sadati M, Taheri Qazvini N, Krishnan R, Park CY, Fredberg JJ (2013) Collective migration and cell jamming *Differentiation* 86:121-125 doi:10.1016/j.diff.2013.02.005
- Saw TB et al. (2017) Topological defects in epithelia govern cell death and extrusion *Nature* 544:212-216 doi:10.1038/nature21718
- Scarpa E, Mayor R (2016) Collective cell migration in development *J Cell Biol* 212:143-155 doi:10.1083/jcb.201508047
- Schwarz US, Soine JR (2015) Traction force microscopy on soft elastic substrates: A guide to recent computational advances *Biochim Biophys Acta* 1853:3095-3104 doi:10.1016/j.bbamcr.2015.05.028
- Sepulveda N, Petitjean L, Cochet O, Grasland-Mongrain E, Silberzan P, Hakim V (2013) Collective cell motion in an epithelial sheet can be quantitatively described by a stochastic interacting particle model *PLoS Comput Biol* 9:e1002944 doi:10.1371/journal.pcbi.1002944
- Sixt M, Engelhardt B, Pausch F, Hallmann R, Wendler O, Sorokin LM (2001) Endothelial cell laminin isoforms, laminins 8 and 10, play decisive roles in T cell recruitment across the blood-brain barrier in experimental autoimmune encephalomyelitis *The Journal of cell biology* 153:933-946 doi:10.1083/jcb.153.5.933
- Song J et al. (2017) Endothelial Basement Membrane Laminin 511 Contributes to Endothelial Junctional Tightness and Thereby Inhibits Leukocyte Transmigration *Cell reports* 18:1256-1269 doi:10.1016/j.celrep.2016.12.092

1 Steinberg MS (2007) Differential adhesion in morphogenesis: a modern view *Curr Opin Genet Dev* 17:281-286
2 doi:10.1016/j.gde.2007.05.002
3 Sunyer R et al. (2016) Collective cell durotaxis emerges from long-range intercellular force transmission
4 *Science* 353:1157-1161 doi:10.1126/science.aaf7119
5 Sutherland HG, Kearns M, Morgan HD, Headley AP, Morris C, Martin DI, Whitelaw E (2000) Reactivation of
6 heritably silenced gene expression in mice *Mamm Genome* 11:347-355
7 Tambe DT et al. (2011) Collective cell guidance by cooperative intercellular forces *Nat Mater* 10:469-475
8 doi:10.1038/NMAT3025
9 Tarle V, Ravasio A, Hakim V, Gov NS (2015) Modeling the finger instability in an expanding cell monolayer
10 *Integr Biol (Camb)* 7:1218-1227 doi:10.1039/c5ib00092k
11 Trappe V, Prasad V, Cipelletti L, Segre PN, Weitz DA (2001) Jamming phase diagram for attractive particles
12 *Nature* 411:772-775 doi:10.1038/35081021
13 Trepats X, Fredberg JJ (2011) Plithotaxis and emergent dynamics in collective cellular migration *Trends in Cell*
14 *Biology* 21:638-646 doi:10.1016/j.tcb.2011.06.006
15 Trepats X, Sahai E (2018) Mesoscale physical principles of collective cell organization *Nature Physics* 14:671-
16 682 doi:10.1038/s41567-018-0194-9
17 Trepats X, Wasserman MR, Angelini TE, Millet E, Weitz DA, Butler JP, Fredberg JJ (2009) Physical forces
18 during collective cell migration *Nature Physics* 5:426-430 doi:10.1038/nphys1269
19 Vishwakarma M, Di Russo J, Probst D, Schwarz US, Das T, Spatz JP (2018) Mechanical interactions among
20 followers determine the emergence of leaders in migrating epithelial cell collectives *Nature*
21 *communications* 9:3469 doi:10.1038/s41467-018-05927-6
22 Wagstaff L, Kolahgar G, Piddini E (2013) Competitive cell interactions in cancer: a cellular tug of war *Trends*
23 *Cell Biol* 23:160-167 doi:10.1016/j.tcb.2012.11.002
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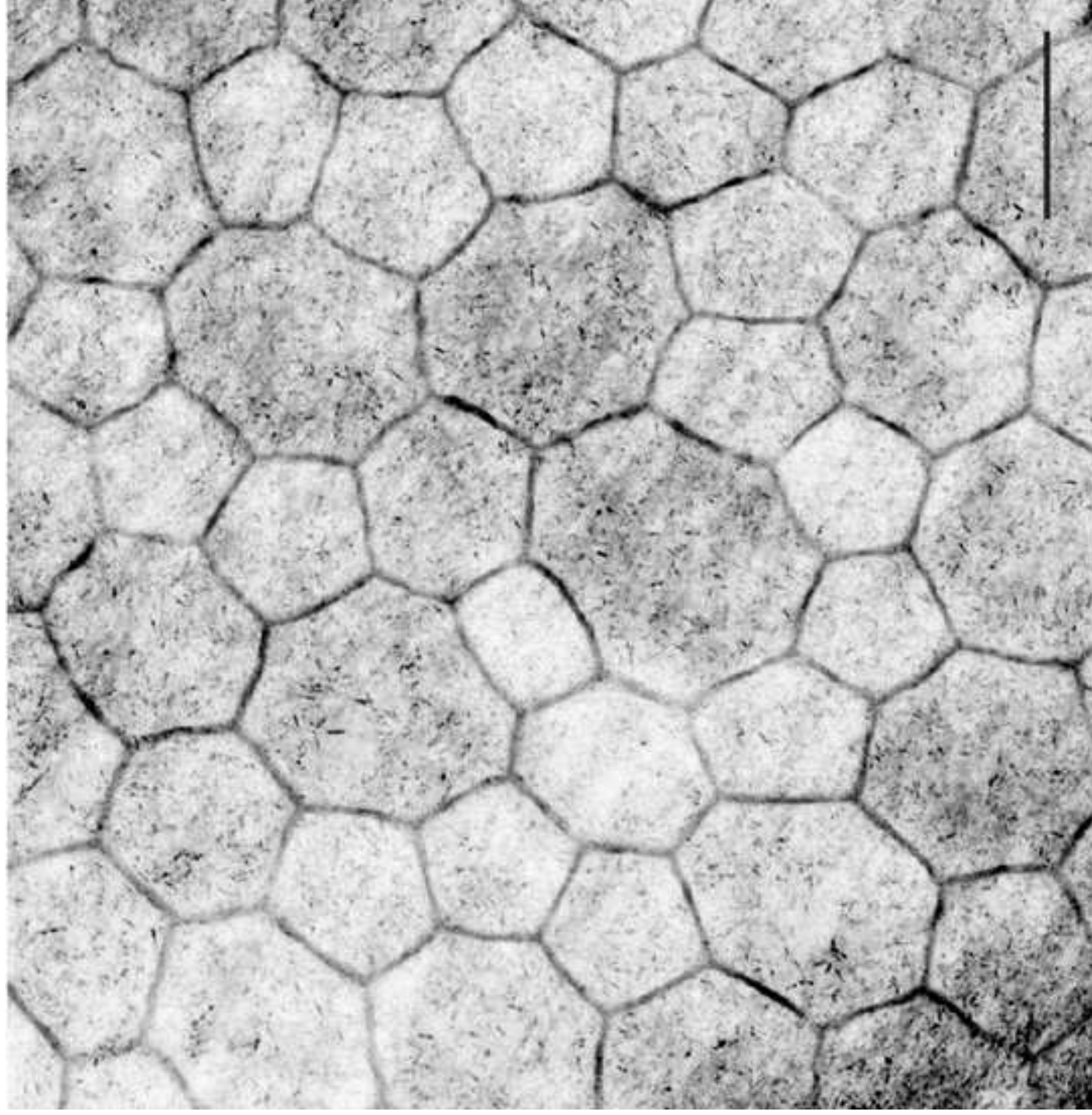


Figure 2

Figure legends.

Figure 1: a) The intercellular stress profile in a confluent epithelial monolayer of canine kidney epithelial cells (MDCK) reveal a rugged stress profile at a given time point. Scale bar is 50µm. b) Cellular heterogeneities can arise from genetic differences or differential regulation of protein expression which are also influenced by external cues such as ECM components. In addition, heterogenous clones in epithelia might differ in their mechanical properties, having different levels of adhesion forces (cell-cell stresses and cell-ECM tractions), thus impacting on physical nature of epithelia.

Figure 2: Immunofluorescent staining of *en-face* preparation of murine retinal pigment epithelium for F-actin reveals highly heterogenous character of this epithelium. To be noted, is the postmitotic nature of these epithelial cells, that excludes correlation of cell size with the cell cycle. Scale bar is 20 µm.